



RESEARCH ARTICLE

Synthesis of prenylated homoisoflavonoids with a coumarin moiety

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Abstract: A series of prenylated homoisoflavonoid-coumarin hybrids were synthesized by developed cascade reactions of coumarin Mannich bases with 3-(dimethylamino)-1-(5-hydroxy-7-methoxy-2,2-dimethyl-3,4-dihydro-2*H*-chromen-6-yl)prop-2-en-1-one. Obtained homoisoflavonoids might have a large potential for further investigations of their bioactivities.

Keywords: coumarins; homoisoflavonoids; inverse electron-demand Diels-Alder reaction.

Introduction

The development of new synthetic routes for the construction of natural product-like small-molecule libraries with structural complexity, diversity, and various physicochemical properties is attracting attention from the scientific community involved in chemical biology and drug discovery. The incorporation of privileged substructural motifs a concept that there exist preferred molecular scaffolds that can provide ligands for diverse receptors has been proven to be an essential tool for the discovery of bioactive molecules [1]. The naturally occurring homoisoflavonoids possess a broad spectrum of biological activities, such as antioxidant, anti-microbial, anti-diabetic, immunomodulatory effects, and protein kinase inhibition [2-4].

The sappanin-type homoisoflavonoids contain chromanone or chromone rings with 3-benzyl substituents. Like other flavonoid counterparts, naturally occurring homoisoflavonoids typically bear hydroxylic, methoxy, and methylenedioxy groups in rings A and B. Isoprenyl and geranyl moieties, methyl groups, and aldehydes have been described as C-linked substituents [5].

Isolated from the bulbs of *Ledebouria Floribunda* prenylated and geranylated flavonoids are very common in certain families of plants. The first examples of prenylated homoisoflavonoids are ledebourin A, B, and C **1a-1c** (Figure 1). These derivatives are the products of triple (acetate, shikimate, and mevalonate) biosynthetic pathways and have been reported as potent antioxidants. The expected activity due to the *ortho*-dihydroxy groups is probably enhanced by other factors introduced by the prenyl and geranyl groups [6].

Results and discussion

It should be noted that a few types of C-prenylated flavonoids have been isolated from natural sources. In most cases, naturally occurring flavonoids contain 2*H*-2,2-dimethylchromene or isopropylbenzofuran fragment, and ring-opening isoprenyl substituent which can be transformed to hydroxy or epoxy derivatives etc. Intramolecular cyclization of *ortho*-hydroxyprenylated flavonoids affords rarely isolated 2,2-dimethylchromane derivatives, which usually undergo oxidation to 2*H*-2,2-dimethylchromene derivatives. For example, a few C-isoprenyl flavonoids **2** [7, 8] as well as their cyclic isomers **3** [7, 9] were isolated from *Epimedium diphilum*, *Bursera leptophloes* (Figure 1).

Direct prenylation of phenols is a challenging problem for the synthesis of prenylated flavonoids. In the case of coumarins and chromones, direct prenylation was achieved using prenyl- or geranyl halides [10-14], 2-methyl-3-butene-2-ol [15], and enzyme-catalyzed prenylation with dimethylallyl diphosphate or geranyl diphosphate [16-18].

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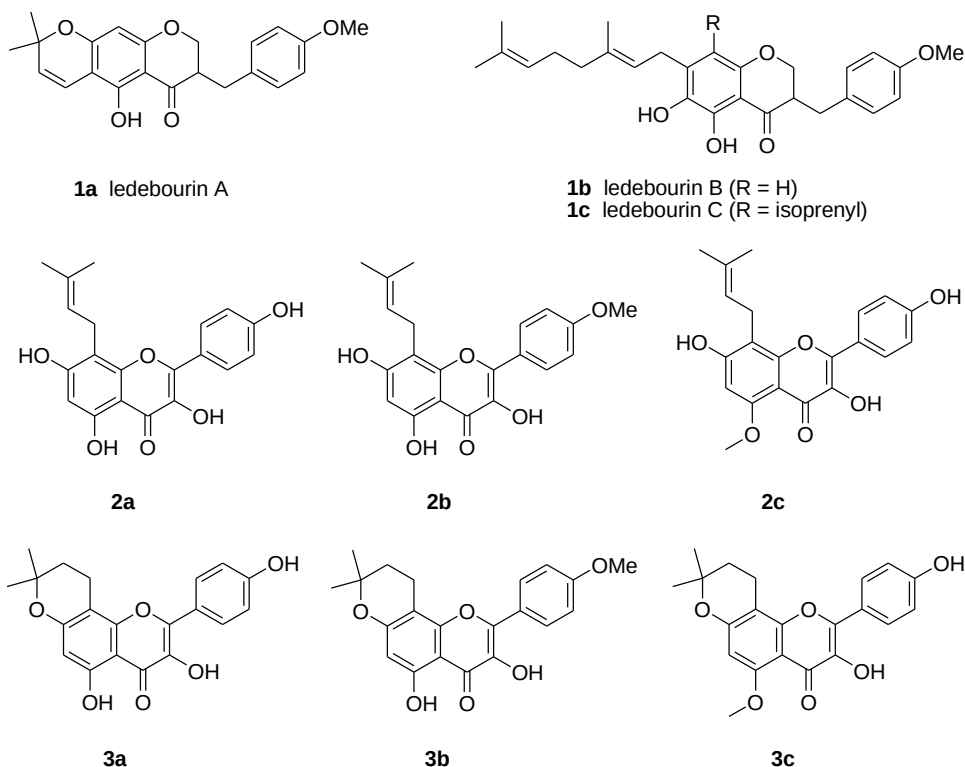
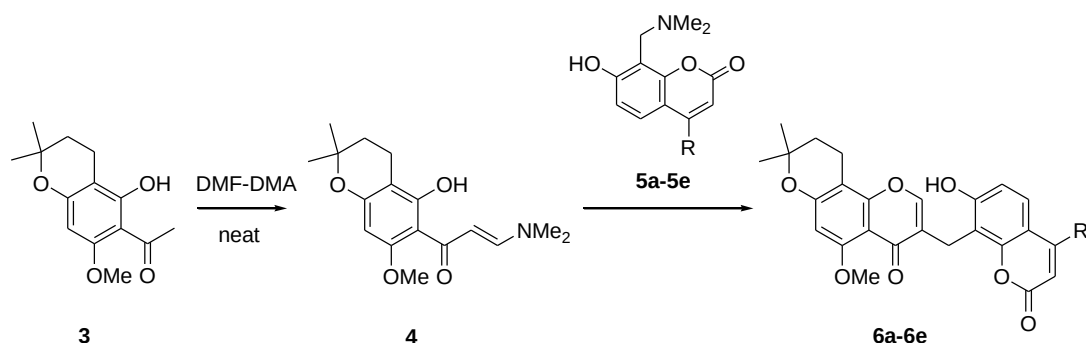


Figure 1. Chemical structures of some prenylated sappanin-type natural homoisoflavonoids and ring-chain isomeric prenylated flavones.



Scheme 1. Synthesis of homoisoflavonoid-coumarin hybrids.

However, in a similar condition, the formation of isoprenyl-substituted compounds and/or 2,2-dimethylchromanes was observed. It should be noted, these reactions had poor selectivity and yield; the formation of O-prenylated, 6,6- or 8,8-bisprenylated compounds was observed using 5,7-dihydroxycoumarins [11, 19] and 5,7-dihydroxychromones [20]; C-3 prenylation of coumarins also was observed [10].

Moreover, this reaction is impossible in the case of A- and B- ring flavonoids due to the low selectivity of prenylation. For the synthesis of A-ring prenylated homoisoflavonoids-coumarin hybrids, we used described one-pot procedure which includes inverse electron-demand Diels-Alder reaction with subsequent intramolecular cascade reactions (Scheme 1) [21].

Started enaminone **4** was synthesized by a reaction of 1-(5-hydroxy-7-methoxy-2,2-dimethylchroman-6-yl)ethan-1-one (**3**) with dimethylformamide dimethyl acetal. 8-Dimethylaminomethyl-7-hydroxycoumarins were synthesized by aminomethylation of 7-hydroxycoumarins with bis(dimethylamino)methane in propanol-2 solution.

The reaction of enaminone **4** with coumarin Mannich bases **5a-5e** was carried out in DMF at reflux for 6 h. The formation of compounds **6a-6e** could be considered as cascade reactions which include: the thermally generated formation of quinone methides **7** from coumarin Mannich bases **5**, inverse electron-demand Diels-Alder reaction of compounds **7** with enaminone **4**, deamination of hemiaminals **8** with the formation of 4*H*-chromeno derivatives **9**, and intramolecular nucleophilic attack of C-4 with the

phenolic group. As result, compounds **6a-6e** were alone products of the reaction of enaminone **4** with 8-dimethylaminomethylcoumarins **5a-5e** (Scheme 2).

Conclusions

As a result, we demonstrated the efficient synthesis of 3-(7-hydroxy-2-oxo-2*H*-chromen-8-ylmethyl)-5-methoxy-8,8-dimethyl-9,10-dihydro-4*H*,8*H*-pyrano[2,3-*f*]chromen-4-ones as isomers of prenylated homoisoflavonoid-coumarin hybrids by the cascade reaction of (2*E*)-3-(dimethylamino)-1-(5-hydroxy-7-methoxy-2,2-dimethyl-3,4-dihydro-2*H*-chromen-6-yl)prop-2-en-1-one with 7-hydroxycoumarin Mannich bases.

Experimental section

^1H and ^{13}C spectra were recorded on Bruker 500 (500/125 MHz) or Bruker 400 (400/100 MHz) spectrometers in CDCl_3 [residual CHCl_3 ($\delta_{\text{H}} = 7.26$ ppm) or CDCl_3 ($\delta_{\text{C}} = 77.16$ ppm) as internal standard] or $\text{DMSO}-d_6$ [residual $\text{SO}(\text{CD}_3)(\text{CD}_2\text{H})$ ($\delta_{\text{H}} = 2.50$ ppm) or $\text{SO}(\text{CD}_3)_2$ ($\delta_{\text{C}} = 39.52$ ppm) as internal standard]. Melting points were determined in open capillary tubes using Buchi B-535 apparatus and were uncorrected. Mass spectra were obtained using an Agilent 1100 spectrometer using APCI (atmospheric-pressure chemical ionization). Elemental analysis was performed on a Vario MICRO cube automated CHNS-analyzer. Column chromatography was performed using Macherey-Nagel Silica 60, 0.04-0.063 mm silica gel.

Compound **5a** was synthesized as reported previously [21].

(2*E*)-3-(Dimethylamino)-1-(5-hydroxy-7-methoxy-2,2-di-methyl-3,4-dihydro-2*H*-chromen-6-yl)prop-2-en-1-one (**4**).

A mixture of 0.75 g (3 mmol) in 2 ml DMF-DMA was heated at reflux for 1 h, cooled, and formed residue was filtered off and washed with MeOH. Compound **4** was recrystallized from MeOH.

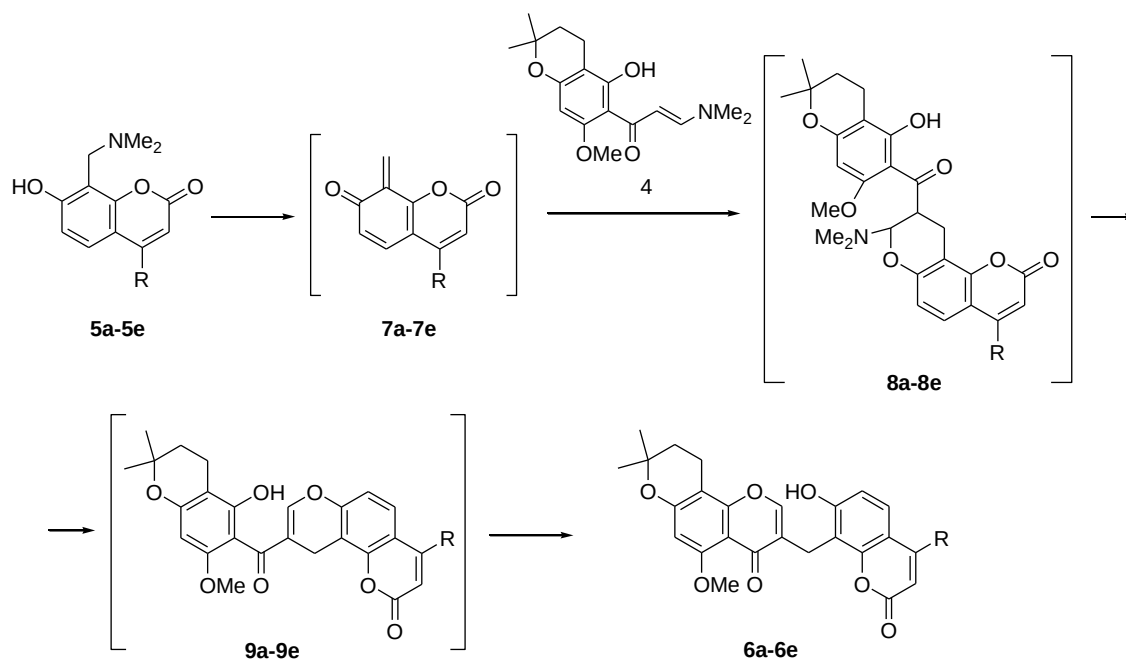
Yield 513 mg, 56%; mp 187-189 °C. ^1H NMR (400 MHz, CDCl_3) δ 15.98 (s, 1H), 7.88 (d, *J* 12.4 Hz, 1H), 6.26 (d, *J* 12.4 Hz, 1H), 5.82 (s, 1H), 3.79 (s, 3H), 3.09 (s, 3H), 2.91 (s, 3H), 2.60 (d, *J* 6.7 Hz, 2H), 1.77 (t, *J* 6.7 Hz, 2H), 1.32 (s, 6H). ^{13}C NMR (125 MHz, CDCl_3) δ 190.2, 165.1, 159.9, 158.4, 154.0, 104.5, 102.0, 97.0, 91.1, 75.4, 55.5, 45.1, 37.3, 32.4, 26.8, 16.4. LC/MS (APCI) *m/z* 306.2 $[\text{M}+\text{H}]^+$. Anal. calcd. for $\text{C}_{17}\text{H}_{23}\text{NO}_4$: C, 66.86; H, 7.59; N, 4.59. Found: C, 66.71; H, 7.48; N, 4.81.

General procedure for the synthesis of 8-dimethylaminomethylcoumarins 5.

To a stirred suspension of coumarins **5c-5e** (2 mmol) in 10 mL of isopropanol was added 0.3 mL (2.2 mmol, 1.1 eq) of bis(*N,N*-dimethylamino)methane at 70 °C. The mixture was heated at 80 °C for 2 h and cooled, and diluted with hexane. The formed residue was filtered off and washed with hexane. The Mannich bases **5** were recrystallized from isopropanol-hexane.

8-[(Dimethylamino)methyl]-7-hydroxy-4-methyl-2*H*-chromen-2-one (**5b**).

Yield 271 mg, 58%; mp 95-97 °C. ^1H NMR (400 MHz, CDCl_3) δ 12.55 (s, 1H), 7.33 (d, *J* 8.7 Hz, 1H), 6.68 (d, *J* 8.7 Hz, 1H), 5.98 (s, 1H), 3.92 (s, 2H), 2.33 (s, 6H), 2.31 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 162.9, 161.1, 153.3,



Scheme 2. Plausible reaction mechanism.

152.3, 124.6, 113.3, 112.0, 110.4, 108.0, 55.1, 44.4, 18. LC/MS (APCI) m/z 234.2 $[M+H]^+$. Anal. calcd. for $C_{13}H_{15}NO_3$: C, 66.94; H, 6.48; N, 6.00. Found: C, 67.31; H, 6.71; N, 6.25.

8-[(Dimethylamino)methyl]-7-hydroxy-4-(methoxymethyl)-2H-chromen-2-one (**5c**).

Yield 448 mg, 85%; mp 126-128 °C. 1H NMR (500 MHz, $CDCl_3$) δ 11.20 (s, 1H), 7.33 (d, J 8.7 Hz, 1H), 6.73 (d, J 8.7 Hz, 1H), 6.30 (s, 1H), 4.56 (s, 2H), 4.00 (s, 2H), 3.48 (s, 3H), 2.38 (s, 6H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 163.0, 161.3, 152.6, 152.3, 123.8, 113.5, 109.6, 108.5, 108.3, 70.4, 59.1, 55.2, 44.5. LC/MS (APCI) m/z 264.0 $[M+H]^+$. Anal. calcd. for $C_{14}H_{17}NO_4$: C, 63.87; H, 6.51; N, 5.32. Found: C, 64.13; H, 6.26; N, 5.44.

8-[(Dimethylamino)methyl]-7-hydroxy-4-isopropyl-2H-chromen-2-one (**5d**).

Yield 392 mg, 75%; mp 130-132 °C. 1H NMR (400 MHz, $CDCl_3$) δ 11.40 (s, 1H), 7.41 (d, J 8.9 Hz, 1H), 6.68 (d, J 8.8 Hz, 1H), 6.02 (s, 1H), 3.93 (s, 2H), 3.17 (hept, J 6.8 Hz, 1H), 2.32 (s, 6H), 1.21 (d, J 6.8 Hz, 6H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 163.0, 162.6, 161.9, 152.6, 124.1, 113.3, 110.7, 108.3, 106.4, 55.2, 44.4, 28.5, 21.9. LC/MS (APCI) m/z 262.2 $[M+H]^+$. Anal. calcd. for $C_{15}H_{19}NO_3$: C, 68.94; H, 7.33; N, 5.36. Found: C, 68.77; H, 7.52; N, 5.21.

4-Cyclopropyl-8-[(dimethylamino)methyl]-7-hydroxy-2H-chromen-2-one (**5e**).

Yield 482 mg, 93%; mp 136-138 °C. 1H NMR (400 MHz, $CDCl_3$) δ 12.57 (s, 1H), 7.67 (d, J 8.8 Hz, 1H), 6.72 (d, J 8.8 Hz, 1H), 5.75 (s, 1H), 3.94 (s, 2H), 2.34 (s, 6H), 2.11-1.88 (m, 1H), 1.16-0.97 (m, 2H), 0.87-0.62 (m, 2H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 162.9, 161.9, 158.7, 152.2, 124.7, 113.3, 112.3, 108.0, 105.4, 55.2, 44.5, 12.0, 8.1. LC/MS (APCI) m/z 260.2 $[M+H]^+$. Anal. calcd. for $C_{15}H_{17}NO_3$: C, 69.48; H, 6.61; N, 5.40. Found: C, 69.73; H, 6.45; N, 5.67.

General procedure for the synthesis of homoisoflavonoids **6**.

A mixture of enaminone **4** (1 mmol) and corresponding Mannich base **5a-e** (1 mmol) in DMF (10 mL) was stirred at reflux for 6 h. The reaction mixture was cooled, the solvent was evaporated, and formed residue was washed with water and purified by recrystallization from DMF-MeOH mixture.

3-[(7-Hydroxy-2-oxo-2H-chromen-8-yl)methyl]-5-methoxy-8,8-dimethyl-9,10-dihydro-4H,8H-pyrano[2,3-f]-chromen-4-one (**6a**).

Yield 326 mg, 75%; mp 263-265 °C. 1H NMR (400 MHz, $DMSO-d_6$) δ 10.67 (s, 1H), 7.92 (d, J 9.4 Hz, 1H), 7.53 (s, 1H), 7.44 (d, J 8.5 Hz, 1H), 6.87 (d, J 8.5 Hz, 1H), 6.32 (s, 1H), 6.20 (d, J 9.4 Hz, 1H), 3.77 (s, 3H), 3.70 (s, 2H), 2.58 (t, J 6.7 Hz, 2H), 1.75 (t, J 6.7 Hz, 2H), 1.27 (s, 6H). ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 175.6, 160.4, 159.2, 158.4, 157.9, 156.4, 153.6, 150.4, 144.8, 127.7, 122.0,

112.9, 111.6, 111.5, 111.2, 107.5, 100.8, 96.8, 75.9, 55.9, 31.0, 26.2, 18.8, 15.7. LC/MS (APCI) m/z 435.2 $[M-H]^-$. Anal. calcd. for $C_{25}H_{22}O_7$: C, 69.12; H, 5.10. Found: C, 69.03; H, 5.38.

3-[(7-Hydroxy-4-methyl-2-oxo-2H-chromen-8-yl)methyl]-5-methoxy-8,8-dimethyl-9,10-dihydro-4H,8H-pyrano[2,3-f]-chromen-4-one (**6b**).

Yield 309 mg, 69%; mp 268-270 °C. 1H NMR (400 MHz, $DMSO-d_6$) δ 10.62 (s, 1H), 7.56-7.48 (m, 2H), 6.88 (d, J 8.8 Hz, 1H), 6.31 (s, 1H), 6.12 (s, 1H), 3.77 (s, 3H), 3.70 (s, 2H), 2.58 (t, J 6.6 Hz, 2H), 2.35 (s, 3H), 1.75 (d, J 6.6 Hz, 2H), 1.27 (s, 6H). ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 175.6, 160.2, 159.0, 158.4, 157.9, 156.4, 153.7, 152.9, 150.3, 124.4, 122.1, 112.6, 112.3, 111.5, 110.1, 107.5, 100.8, 96.8, 75.9, 55.9, 31.0, 26.2, 18.9, 18.2, 15.7. LC/MS (APCI) m/z 449.0 $[M+H]^+$. Anal. calcd. for $C_{26}H_{24}O_7$: C, 69.63; H, 5.39. Found: C, 69.84; H, 5.53.

3-[[7-Hydroxy-4-(methoxymethyl)-2-oxo-2H-chromen-8-yl)methyl]-5-methoxy-8,8-dimethyl-9,10-dihydro-4H,8H-pyrano[2,3-f]-chromen-4-one (**6c**).

Yield 388 mg, 81%; mp 261-263 °C. 1H NMR (400 MHz, $DMSO-d_6$) δ 10.66 (s, 1H), 7.54 (s, 1H), 7.46 (d, J 8.7 Hz, 1H), 6.87 (d, J 8.7 Hz, 1H), 6.33 (s, 1H), 6.19 (s, 1H), 4.62 (s, 2H), 3.77 (s, 3H), 3.71 (s, 2H), 3.41 (s, 3H), 2.60 (t, J 6.7 Hz, 2H), 1.76 (t, J 6.7 Hz, 2H), 1.28 (s, 6H). ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 175.6, 160.3, 159.0, 158.4, 157.9, 156.4, 153.1, 152.8, 150.4, 123.7, 122.0, 112.7, 111.7, 109.9, 107.8, 107.5, 100.9, 96.9, 76.0, 69.5, 58.4, 55.9, 31.0, 26.2, 19.0, 15.7. LC/MS (APCI) m/z 479.0 $[M+H]^+$. Anal. calcd. for $C_{27}H_{26}O_8$: C, 67.77; H, 5.48. Found: C, 67.53; H, 5.65.

3-[(7-Hydroxy-4-isopropyl-2-oxo-2H-chromen-8-yl)methyl]-5-methoxy-8,8-dimethyl-9,10-dihydro-4H,8H-pyrano[2,3-f]-chromen-4-one (**6d**).

Yield 372 mg, 78%; mp 246-248 °C. 1H NMR (400 MHz, $DMSO-d_6$) δ 10.61 (s, 1H), 7.64 (d, J 7.2 Hz, 1H), 7.55 (s, 1H), 6.90 (d, J 8.7 Hz, 1H), 6.34 (s, 1H), 6.09 (s, 1H), 3.78 (s, 3H), 3.71 (s, 2H), 3.34-3.26 (m, 1H), 2.61 (d, J 6.6 Hz, 1H), 1.77 (d, J 6.6 Hz, 3H), 1.28 (s, 6H), 1.23 (d, J 6.7 Hz, 6H). ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 175.6, 162.9, 160.7, 158.8, 158.4, 158.0, 156.4, 153.2, 150.4, 123.9, 122.1, 112.8, 111.9, 111.0, 107.5, 106.3, 100.8, 96.9, 75.9, 55.9, 31.0, 28.0, 26.2, 21.7, 19.0, 15.7. LC/MS (APCI) m/z 477.0 $[M+H]^+$. Anal. calcd. for $C_{28}H_{28}O_7$: C, 70.58; H, 5.92. Found: C, 70.36; H, 6.17.

3-[(4-Cyclopropyl-7-hydroxy-2-oxo-2H-chromen-8-yl)methyl]-5-methoxy-8,8-dimethyl-9,10-dihydro-4H,8H-pyrano[2,3-f]-chromen-4-one (**6e**).

Yield 266 mg, 56%; mp 244-246 °C. 1H NMR (400 MHz, $DMSO-d_6$) δ 10.64 (s, 1H), 7.81 (d, J 8.8 Hz, 1H), 7.53 (s, 1H), 6.91 (d, J 8.8 Hz, 1H), 6.31 (s, 1H), 5.81 (s, 1H), 3.77 (s, 3H), 3.71 (s, 2H), 2.56 (t, J 6.6 Hz, 2H), 2.27-2.12 (m, 1H), 1.73 (d, J 6.6 Hz, 2H), 1.25 (s, 6H), 1.13-1.00 (m, 2H), 0.91-0.77 (m, 2H). ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 175.6, 160.7, 159.0, 158.9, 158.4, 157.9, 156.4, 152.8, 150.4, 124.3, 122.1, 112.7, 112.5, 111.6, 107.5, 104.5,

100.8, 96.8, 75.9, 55.9, 31.0, 26.2, 19.0, 15.7, 11.5, 8.6. LC/MS (APCI) m/z 475.0 $[M+H]^+$. Anal. calcd. for $C_{28}H_{26}O_7$: C, 70.87; H, 5.52. Found: C, 70.63; H, 5.76.

Notes

The authors declare no conflict of interest.

References

- An, H.; Eum, S.-J.; Koh, M.; Lee, S.K.; Park, S.B. Diversity-Oriented Synthesis of Privileged Benzopyranyl Heterocycles from *s*-cis-Enones. *J. Org. Chem.* **2008**, *73*, 1752-1761.
- Siddaiah, V.; Rao, C.V.; Venkateswarlu, S.; Krishnaraju, A.V.; Subbaraju, G.V. Synthesis, stereochemical assignments, and biological activities of homoisoflavonoids. *Bioorg. Med. Chem.* **2006**, *14*, 2545-2551.
- Abegaz, B.M.; Mutanyatta-Comar, J.; Nindi, M. Naturally Occurring Homoisoflavonoids: Phytochemistry, Biological Activities and Synthesis. *Nat. Prod. Commun.* **2007**, *2*, 475-498.
- Kumar, V.; Nayak, S.K. Homoisoflavonoids: isolation, chemical synthesis strategies and biological activities. *J. Pharm. Sci. Res.* **2020**, *12*, 1046-1055.
- Castelli, M.V.; López, S.N. in *Studies in Natural Products Chemistry*, edited by Atta-ur-Rahman (Elsevier, 2017), Vol. 54, pp. 315-354.
- Abegaz, B.M.; Kinf, H.H. Naturally Occurring Homoisoflavonoids: Phytochemistry, Biological Activities, and Synthesis (Part II). *Nat. Prod. Commun.* **2019**, *14*, 1934578X19845813.
- Souza, M.P.; Machado, M.I.L.; Braz-Filho, R. Six flavonoids from *Bursera leptophloeos*. *Phytochemistry* **1989**, *28*, 2467-2470.
- Mizuno, M.; Iinuma, M.; Tanaka, T.; Sakakibara, N.; Fujikawa, T.; Hanioka, S.; Ishida, Y.; Liu, X.-S.; Murata, H. Flavonol glycosides in the roots of *Epimedium diphyllum*. *Phytochemistry* **1988**, *27*, 3645-3647.
- Guo, B.-L.; Li, W.-K.; Yu, J.-G.; Xiao, P.-G. Brevicornin, a flavonol from *Epimedium brevicornum*. *Phytochemistry* **1996**, *41*, 991-992.
- Crombie, L.; Jones, R.C.F.; Palmer, C.J. Synthesis of mameins and surangin a. *Tetrahedron Lett.* **1985**, *26*, 2929-2932.
- Crombie, L.; Jones, R.C.F.; Palmer, C.J. Synthesis of the insecticidal 1'-acetoxy-mameins and surangin b. *Tetrahedron Lett.* **1985**, *26*, 2933-2936.
- Jetter, M.M.; Heindel, N.D.; Laskin, J.D. Novel syntheses of dihydroxanthyletin and dihydroseselin derivatives. *J. Heterocycl. Chem.* **1990**, *27*, 995-997.
- Daskiewicz, J.-B.; Depeint, F.; Viorner, L.; Bayet, C.; Comte-Sarrasin, G.; Comte, G.; Gee, J.M.; Johnson, I.T.; Ndjoko, K.; Hostettmann, K.; Barron, D. Effects of Flavonoids on Cell Proliferation and Caspase Activation in a Human Colonic Cell Line HT29: An SAR Study. *J. Med. Chem.* **2005**, *48*, 2790-2804.
- Garazd, Y.L.; Garazd, M.M.; Khilya, V.P. Modified Coumarins. 16. Cyclohexane-Annelated Analogs of Pyranocoumarins. *Chem. Nat. Compd.* **2005**, *41*, 388-395.
- Lee, J.H.; Bang, H.B.; Han, S.Y.; Jun, J.-G. An efficient synthesis of (+)-decursinol from umbelliferone. *Tetrahedron Lett.* **2007**, *48*, 2889-2892.
- Chen, R.; Liu, X.; Zou, J.; Yin, Y.; Ou, B.; Li, J.; Wang, R.; Xie, D.; Zhang, P.; Dai, J. Regio- and Stereospecific Prenylation of Flavonoids by *Sophora flavescens* Prenyltransferase. *Adv. Synth. Catal.* **2013**, *355*, 1817-1828.
- Li, J.; Chen, R.; Wang, R.; Liu, X.; Xie, D.; Zou, J.; Dai, J. GuA6DT, a Regiospecific Prenyltransferase from *Glycyrrhiza uralensis*, Catalyzes the 6-Prenylation of Flavones. *ChemBioChem* **2014**, *15*, 1673-1681.
- Xu, Y.; Li, D.; Tan, G.; Zhang, Y.; Li, Z.; Xu, K.; Li, S.-M.; Yu, X.-A. Single Amino Acid Switch Alters the Prenyl Donor Specificity of a Fungal Aromatic Prenyltransferase toward Biflavonoids. *Org. Lett.* **2021**, *23*, 497-502.
- Crombie, L.; Jones, R.C.F.; Palmer, C.J. Synthesis of the Mamea coumarins. Part 1. The coumarins of the mamea A, B, and C series. *J. Chem. Soc., Perkin Trans. 1* **1987**, 317-331.
- Neves, M.P.; Cidade, H.; Pinto, M.; Silva, A.M.S.; Gales, L.; Damas, A.M.; Lima, R.T.; Vasconcelos, M.H.; Nascimento, M. d. S.J. Prenylated derivatives of baicalein and 3,7-dihydroxyflavone: Synthesis and study of their effects on tumor cell lines growth, cell cycle and apoptosis. *Eur. J. Med. Chem.* **2011**, *46*, 2562-2574.
- Mrug, G.P.; Myshko, N.V.; Bondarenko, S.P.; Sviripa, V.M.; Frasinuk, M.S. One-Pot Synthesis of B-Ring *ortho*-Hydroxylated Sappanin-type Homoisoflavonoids. *J. Org. Chem.* **2019**, *84*, 7138-7147.

Синтез пренільованих гомоізофлавоноїдів із кумариновим фрагментом

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Резюме: Синтезовано ряд пренільованих гомоізофлавоноїд-кумаринових гібридів шляхом каскадних реакцій кумаринових основ Манніха з 3-(диметиламіно)-1-(5-гідрокси-7-метокси-2,2-диметил-3,4-дигідро-2H-хромен-6-іл)проп-2-ен-1-оном. Отримані гомоізофлавоноїди можуть мати великий потенціал для подальших досліджень їх біологічної активності.

Ключові слова: кумарини; гомоізофлавоноїди; обернена за електронними вимогами реакція Дільса-Альдера.